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(54) Title: CAROTENOID KETOLASE GENES AND GENE PRODUCTS, PRODUCTION OF KETOCAROTENOID AND METH- ODS OF MODIFYING CAROTENOID USING THE GENES (57) Abstract A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3, or has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, as well as vectors and host cells containing them. Methods of use of the nucleic acid sequences to produce ketocarotenoid in host cells and methods of use of the nucleic acid sequences to modify the production of carotenoids in a host cell are included.		

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**CAROTENOID KETOLASE GENES AND GENE PRODUCTS,
PRODUCTION OF KETOCAROTENOIDS AND METHODS OF
MODIFYING CAROTENOIDS USING THE GENES**

BACKGROUND OF THE INVENTION

5 Carotenoids are widely distributed natural pigments that are responsible for many of the yellow, orange and red colors seen in living organisms. They have important commercial uses as coloring agents in the food industry, as feed and food additives, in cosmetics and as provitamin A precursors.

The plant species *Adonis aestivalis* produces flowers with petals that are deep
10 red in color and nearly black at the base of the petals due to the accumulation of ketocarotenoid and other carotenoid pigments (Neamtu et al., *Rev. Roum. Biochim.* 6:157, 1969). This pattern of carotenoid accumulation accounts for the common name of some varieties of this species: summer pheasant's eye.

Among the carotenoids identified in the petals of the red petal varieties of these
15 various species is the ketocarotenoid astaxanthin (3,3'-dihydroxy-4,4'-diketo-b,b-carotene; see Figure 1). Various other ketocarotenoids (see Figure 1) including 3-hydroxyechinenone (3-hydroxy-4-keto-b,b-carotene), adonirubin (3-hydroxy-4,4'-diketo-b,b-carotene) adonixanthin (3,3'-dihydroxy-4-keto-b,b-carotene) and isozeaxanthin (4,4'-dihydroxy-b,b-carotene; see T.W. Goodwin, *The Biochemistry of the Carotenoids*,
20 vol I. Plants, 2nd edition, 1980, page 147) have also been reported. The latter compound is consistent with speculation that the 4-hydroxy may be an intermediate in the formation of the 4-keto group.

SUMMARY OF THE INVENTION

There is appreciable interest in the biological production of carotenoids, in
25 particular the orange-colored ketocarotenoids such as astaxanthin and canthaxanthin (Figure 1), and in the modification of carotenoid composition. For this reason, an *A. aestivalis* flower cDNA library was constructed and screened for cDNAs encoding enzymes (hereinafter referred to as "ketolases" although the specific biochemical activity has not yet been established) involved in the conversion of b-carotene into
30 orange compounds with absorption properties similar to those exhibited by common ketocarotenoids such as canthaxanthin (Figure 1). Two distinctly different *Adonis aestivalis* cDNAs were obtained from among a number of cDNAs that were selected on this basis.

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Thus, a first aspect of the present invention is a purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3.

5 The invention also includes a purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and having the amino acid sequence of SEQ ID NO: 2 or 4.

The invention also includes vectors which comprise any portion of the nucleic acid sequences listed above, and host cells transformed with such vectors.

10 Another aspect of the present invention is a method of producing a ketocarotenoid in a host cell, the method comprising

inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked
15 to a promoter; and

expressing the heterologous nucleic acid sequence, thereby producing the ketolase enzyme.

20 Another subject of the present invention is a method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method comprising

inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous
25 nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

BRIEF DESCRIPTION OF THE DRAWINGS

30 A more complete appreciation of the invention and many of the attendant advantages thereof will be readily obtained as the same becomes better understood by

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reference to the following detailed description when considered in connection with the accompanying drawings.

Figure 1 illustrates structures and biochemical routes leading from b-carotene to various of the ketocarotenoids referred to in the text. Conversion of β -carotene to astaxanthin by a hydroxylase enzyme (Hy) and a ketolase enzyme (keto) could proceed via any one or all of several possible routes depending on the order of the reactions.

Figure 2 illustrates the *beta* ring structure of b-carotene and various modifications of this parent ring that might be produced through the action of the products of the *A. aestivalis* ketolase cDNAs. Also shown is the structure of the *epsilon* ring, not found to be a substrate for the *A. aestivalis* ketolases and present in carotenoids such as d-carotene, e-carotene, a-carotene and lutein.

Figure 3 illustrate results obtained with TLC (thin layer chromatography) separation of carotenoid pigments extracted from *E. coli* cultures, previously engineered to produce b-carotene, but that now also contain the *A. aestivalis* ketolase cDNAs and/or other introduced genes and cDNAs. The Figure indicates the empty plasmid vector pBluescript SK- (SK-), the *Adonis aestivalis* ketolase 1 cDNA in this plasmid vector (Ad keto1), the *Haematococcus pluvialis* ketolase cDNA in this plasmid vector Hp keto), or the Arabidopsis β -carotene hydroxylase cDNA (At Ohase). Bands that were orange in color are shown here with a darker fill than those with a yellow color. Identities of various bands are indicated to the right of the band.

Figure 4 illustrates the absorption spectrum of one of the orange carotenoids produced from b-carotene via the action of the *Adonis* ketolases and makes clear the similarity of the spectrum to that of canthaxanthin. Absorption spectra (in acetone) of β -carotene, canthaxanthin and an unknown orange product (orange band #1; the lower orange band in the first lane of Figure 3) extracted from cultures after introduction of the *Adonis aestivalis* keto1 cDNA (SEQ ID NO: 1) in cells of *E. coli* that otherwise produce and accumulate β -carotene. The absorption spectrum of the unknown resembles that of canthaxanthin but the compound migrates to a position below echinenone on RP18

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TLC plates developed with a mobile phase of methanol:acetone (1:1 by volume). The absorption spectrum of orange band #2 also is similar to that of canthaxanthin but it migrates more rapidly than canthaxanthin indicating that it is probably a more polar compound.

- 5 Figure 5 shows SEQ ID NO: 5 (the sequence shown in this Figure includes SEQ ID NO: 1 and also includes some of the flanking DNA from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector, which sequences are shown in bold).

- Figure 6 shows SEQ ID NO: 6 (the sequence shown in this Figure includes SEQ ID NO: 2 and also includes a translation of amino acids resulting from the adaptor DNA and
10 the multiple cloning site (MCS) of the library cloning vector and the start codon from the plasmid vector pTrcHis, which sequences are shown in bold and capitalized).

Figure 7 shows SEQ ID NO: 7 (the sequence shown in this Figure includes SEQ ID NO: 3 and also includes some of the flanking DNA from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector, which sequences are shown in bold).

- 15 Figure 8 shows SEQ ID NO: 8 (the sequence shown in this Figure includes SEQ ID NO: 4 and also includes a translation of amino acids resulting from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector and the start codon from the plasmid vector, which sequences are shown in bold and capitalized).

- Figure 9 shows a "Gap" alignment of the two Adonis ketolase sequences of the
20 invention. A truncated version of SEQ ID NO: 1 is shown in this Figure for comparative purposes, and is designated SEQ ID NO: 9. The percentage identity was calculated to be 91.107.

Figure 10 shows a "Gap" alignment of SEQ ID NO: 2 and 4. The following results were found:

- | | | |
|----|------------------|--------------------------|
| 25 | Gap weight: 12 | average match: 2.912 |
| | Length weight: 4 | average mismatch: -2.003 |

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Quality: 1440 length: 307
 Ratio: 4.691 gaps: 0
 percent similarity: 92.182 percent identity: 90.228

Figure 11 shows a comparison between SEQ ID NO: 2 and the *Arabidopsis thaliana* β -carotene hydroxylase enzyme (GenBank U58919) (SEQ ID NO: 10).

Figure 12A shows gDNA (SEQ ID NO: 11) immediately upstream of the cDNA of SEQ ID NO: 3. The sequence was obtained from a PCR product generated using the GenomeWalker kit of Clontech Laboratories, Inc. (1020 East Meadow Circle, Palo Alto, CA 94303-4230) and nested primers specific to the ketolases of *Adonis aestivalis* (cagaatcgggtctgttctattagtcttcc (SEQ ID NO: 17) and caatttgaggaatatcaaggttcctgttctc (SEQ ID NO: 18)). The termination codon upstream of and in-frame with initiation codon (TAA at positions 204-206) is shown in bold. Initiation codon (ATG) is also shown in bold.

Figure 12B (SEQ ID NO: 12) indicates that the full length polypeptide of SEQ ID NO: 4 begins with the amino acids MAA (shown in bold) immediately preceding the ketolase sequence shown in Figure 8. A similar MAA amino acid sequence immediately preceding SEQ ID NO: 1 is also expected.

Figure 13 shows an alignment of SEQ ID NO: 2, SEQ ID NO: 12, an *Arabidopsis* β -carotene hydroxylase enzyme (predicted product of GenBank U58919) (SEQ ID NO: 13), a putative second *Arabidopsis* hydroxylase predicted by genomic DNA sequence (GenBank AB025606; the exon/intron junctions were chosen with reference to the product of the *Arabidopsis* β -carotene hydroxylase cDNA u58919) (SEQ ID NO: 14), and two *Capsicum annuum* β -carotene hydroxylases (predicted products of GenBank Y09722 and Y09225) (SEQ ID NO: 15 and 16).

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed to a purified nucleic acid sequence which

encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3.

The invention also includes a purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and having the amino acid sequence of SEQ
5 ID NO: 2 or 4.

Two different but closely-related nucleic acids have been isolated. The sequences of the longest example of each are presented herein. Sequencing which has subsequently been conducted of upstream genomic DNA indicates that SEQ ID NO: 3 lacks bases encoding the first three amino acids (MAA; see Figure 12). Likely,
10 this is also the case for SEQ ID NO: 1, but the upstream genomic sequences have not yet been obtained for this nucleic acid.

The two different Adonis ketolases denoted in SEQ ID NO: 1 and 3 are similar in sequence, sharing about 91% identity, as determined by the Gap program discussed below (see Figure 9). The predicted amino acid sequences of the enzymes denoted in
15 SEQ ID NO: 2 and 4 share about 92% similarity and about 90% identity, also as determined by the Gap program (see Figure 10).

Therefore, it is clear that certain modifications of SEQ ID NO: 1 or 3 or SEQ ID NO: 2 or 4 can take place without destroying the activity of the enzyme. Note also that certain truncated versions of the cDNAs of SEQ ID NO: 1 or 3 were found to be
20 functional (i.e., these cDNAs retained the property of causing the conversion of β -carotene to orange compounds). Also, the Arabidopsis β -carotene hydroxylase (GenBank U58919), aligned with the ketolase SEQ ID NO: 2 in Figure 11, retains catalytic function when truncated to yield a polypeptide that lacks the first 129 amino acids (Sun et al., 1996). From the alignment in Figure 11, therefore, this would suggest
25 that the two ketolases of the invention retain catalytic activity after truncation to remove bases encoding the first 132 amino acids.

Thus, the present invention is intended to include those ketolase nucleic acid and amino acid sequences in which substitutions, deletions, additions or other modifications have taken place, as compared to SEQ ID NO: 1 or 3 or SEQ ID NO: 2
30 or 4, without destroying the activity of the ketolase enzyme. Preferably, the substitutions, deletions, additions or other modifications take place at those positions which already show dissimilarity between the present sequences. For SEQ ID NO: 1,

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as shown in Figure 9, these positions are as follows: positions 7, 20, 23, 35, 53, 63, 65, 67, 76, 78, 85, 86, 91, 107, 109-111, 135, 140, 144, 146, 160, 168, 217, 219, 241, 249, 254, 256, 271, 291, 296, 349, 389, 400, 406, 431, 448, 449, 460, 471, 499, 530, 589, 619, 643, 653, 654, 667, 679, 709, 731, 742, 784, 787, 836, 871, 883, 896, 911, 919, 928, 930, 939, 943, 967, 969, 978, 979, 982, 988, 995, 1005, 1006, 1012-1014, 1017, 1019-1021, 1023, 1025, 1049, 1050, 1054, 1060-1068, 1070-1073, 1075, 1094, 1100, 1101, 1106, 1107, 1109 and 1111-1176. For SEQ ID NO: 3, as shown in Figure 9, these positions are as follows: positions 7, 20, 23, 35, 53, 63, 65, 67, 76, 78, 85, 86, 91, 107, 109-111, 135, 140, 144, 146, 160, 168, 217, 219, 241, 249, 254, 256, 271, 291, 296, 349, 389, 400, 406, 431, 448, 449, 460, 471, 499, 530, 589, 619, 643, 653, 654, 667, 679, 709, 731, 742, 784, 787, 836, 871, 883, 896, 911, 919, 928, 930, 939, 943, 966, 967, 970, 979, 980, 983, 989, 996, 1006, 1007, 1013-1015, 1018, 1020-1022, 1024, 1026, 1050, 1051, 1055, 1062-1065, 1067, 1086, 1092, 1093, 1098, 1099, 1101 and 1103-1112.

For SEQ ID NO: 2 and 4, as shown in Figure 10, the following amino acids can be substituted or deleted, or additions or other modifications can be made, without destroying the activity of the ketolase enzyme: positions 7, 8, 12, 18, 21, 22, 25, 26, 36, 37, 45, 47-49, 56, 73, 83, 85, 97, 99, 130, 144, 150, 157, 166, 218, 244, 279, 299 and 304. Therefore, the present invention also intends to cover amino acid sequences where such changes have been made.

In each case, nucleic acid and amino acid sequence similarity and identity is measured using sequence analysis software, for example, the Sequence Analysis, Gap, or BestFit software packages of the Genetics Computer Group (University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wisconsin 53705), MEGAlign (DNASTar, Inc., 1228 S. Park St., Madison, Wisconsin 53715), or MacVector (Oxford Molecular Group, 2105 S. Bascom Avenue, Suite 200, Campbell, California 95008). Such software uses algorithms to match similar sequences by assigning degrees of identity to various substitutions, deletions, and other modifications, and includes detailed instructions as to useful parameters, etc., such that those of routine skill in the art can easily compare sequence similarities and identities. An example of a useful algorithm in this regard is the algorithm of Needleman and Wunsch, which is used in the Gap program discussed above. This program finds the alignment of two complete

sequences that maximizes the number of matches and minimizes the number of gaps. Another useful algorithm is the algorithm of Smith and Waterman, which is used in the BestFit program discussed above. This program creates an optimal alignment of the best segment of similarity between two sequences. Optimal alignments are found by
5 inserting gaps to maximize the number of matches using the local homology algorithm of Smith and Waterman.

Conservative (i.e. similar) substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine and leucine; aspartic acid, glutamic acid, asparagine and glutamine; serine and threonine; lysine and arginine; and
10 phenylalanine and tyrosine. Substitutions may also be made on the basis of conserved hydrophobicity or hydrophilicity (see Kyte and Doolittle, *J. Mol. Biol.* **157**: 105-132 (1982)), or on the basis of the ability to assume similar polypeptide secondary structure (see Chou and Fasman, *Adv. Enzymol.* **47**: 45-148 (1978)).

If comparison is made between nucleotide sequences, preferably the length of
15 comparison sequences is at least 50 nucleotides, more preferably at least 60 nucleotides, at least 75 nucleotides or at least 100 nucleotides. It is most preferred if comparison is made between the nucleic acid sequences encoding the enzyme coding regions necessary for enzyme activity. If comparison is made between amino acid sequences, preferably the length of comparison is at least 20 amino acids, more
20 preferably at least 30 amino acids, at least 40 amino acids or at least 50 amino acids. It is most preferred if comparison is made between the amino acid sequences in the enzyme coding regions necessary for enzyme activity.

While the two different Adonis ketolase enzymes of the present invention are similar in sequence, previously-described bacterial (Misawa et al., 1995), cyanobacterial
25 (Fernandez-Gonzalez et al., 1997), and green algal (*Haematococcus pluvialis*; Lotan et al., 1995; Kajiwara et al., 1995) β -carotene ketolase enzymes bear little resemblance to the Adonis ketolases, although certain histidine motifs and features of the predicted secondary structure are common to the polypeptides predicted by both groups (Cunningham and Gantt, 1998).

30 The present invention also includes vectors containing the nucleic acids of the invention. Suitable vectors according to the present invention comprise a gene encoding a ketolase enzyme as described above, wherein the gene is operably linked

to a suitable promoter. Suitable promoters for the vector can be constructed using techniques well known in the art (see, for example, Sambrook et al., Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York, 1991). Suitable vectors for eukaryotic expression in plants are described in Fray et al., (1995; *Plant J.* 8:693-701) and Misawa et al, (1994; *Plant J.* 6:481-489). Suitable vectors for prokaryotic expression include pACYC184, pUC119, and pBR322 (available from New England BioLabs, Beverly, MA) and pTrcHis (Invitrogen) and pET28 (Novagen) and derivatives thereof. The vectors of the present invention can additionally contain regulatory elements such as promoters, repressors, selectable markers such as antibiotic resistance genes, etc., the construction of which is very well known in the art.

The genes encoding the ketolase enzymes as described above, when cloned into a suitable expression vector, can be used to overexpress these enzymes in a host cell expression system or to inhibit the expression of these enzymes. For example, a vector containing a gene of the invention may be used to increase the amount of ketocarotenoids in an organism and thereby alter the nutritional or commercial value or pharmacology of the organism. A vector containing a gene of the invention may also be used to modify the carotenoid production in an organism.

Therefore, the present invention includes a method of producing a ketocarotenoid in a host cell, the method comprising

inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.

The present invention also includes a method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method comprising inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having

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ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to
5 modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

The term "modifying the production" means that the amount of carotenoids produced can be enhanced, reduced, or left the same, as compared to an untransformed host cell. In accordance with one embodiment of the present invention,
10 the make-up of the carotenoids (i.e., the type of carotenoids produced) is changed *vis a vis* each other, and this change in make-up may result in either a net gain, net loss, or no net change in the amount of carotenoids produced in the cell. In accordance with another embodiment of the present invention, the production or the biochemical activity of the carotenoids (or the enzymes which catalyze their formation) is enhanced by the
15 insertion of the ketolase enzyme-encoding nucleic acid. In yet another embodiment of the invention, the production or the biochemical activity of the carotenoids (or the enzymes which catalyze their formation) may be reduced or inhibited by a number of different approaches available to those skilled in the art, including but not limited to such methodologies or approaches as anti-sense (e.g., Gray et al. (1992), *Plant Mol. Biol.* 19:69-87), ribozymes (e.g., Wegener et al (1994) *Mol. Gen. Genet.* 1994 Nov
20 15;245(4):465-470), co-suppression (e.g. Fray et al. (1993) *Plant Mol. Biol.* 22:589-602), targeted disruption of the gene (e.g., Schaefer et al. *Plant J.* 11:1195-1206, 1997), intracellular antibodies (e.g., see Rondon et al. (1997) *Annu. Rev. Microbiol.* 51:257-283) or whatever other approaches rely on the knowledge or
25 availability of the nucleic acid sequences of the invention, or the enzymes encoded thereby.

Host systems according to the present invention preferably comprise any organism which is capable of producing carotenoids, or which already produces carotenoids. Such organisms include plants, algae, certain bacteria, cyanobacteria and
30 other photosynthetic bacteria. Transformation of these hosts with vectors according to the present invention can be done using standard techniques. See, for example, Sambrook et al., Molecular Cloning A Laboratory Manual, Cold Spring Harbor

Laboratory, Cold Spring Harbor, NY, 1989; Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York, 1991.

Alternatively, transgenic organisms can be constructed which include the nucleic acid sequences of the present invention. The incorporation of these sequences can allow the controlling of carotenoid biosynthesis, content, or composition in the host cell. These transgenic systems can be constructed to incorporate sequences which allow for the overexpression of the various nucleic acid sequences of the present invention. Transgenic systems can also be constructed which allow for the underexpression of the various nucleic acid sequences of the present invention. Such systems may contain anti-sense expression of the nucleic acid sequences of the present invention. Such anti-sense expression would result in the accumulation of the substrates of the enzyme encoded by the sense strand.

Having generally described this invention, a further understanding can be obtained by reference to certain specific examples which are provided herein for purposes of illustration only and are not intended to be limiting unless otherwise specified.

EXAMPLE 1

Isolation of plant cDNAs that convert b-carotene into compounds with ketocarotenoid-like spectra

A flower cDNA library from the plant *Adonis aestivalis* was introduced into a strain of *Escherichia coli* engineered to accumulate the yellow carotenoid pigment β -carotene (see Cunningham et al., *Plant Cell* 8:1613-26, 1996). This strain of *E. coli* normally forms yellow colonies when cultures are spread on a solid agar growth medium. Ketocarotenoids that are derived from b-carotene, such as echinenone and canthaxanthin (Figure 1), are, in contrast, orange to orange-red in color. Colonies that were orange rather than yellow in color were visually selected, and the DNA sequences of the *Adonis aestivalis* cDNAs within the plasmid vectors contained in these colonies were ascertained. Two distinct cDNAs were obtained from analysis of cDNA inserts in plasmids obtained from approximately 10 selected colonies. The DNA sequences of these two ketolase cDNAs are presented herein.

The products produced by the ketolases of the invention which have been

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expressed in a β -carotene-accumulating strain of *Escherichia coli* have not yet been identified. As many as 5 or 6 different colored bands, in addition to the substrate β -carotene, may readily be discerned by C_{18} TLC separation (see Figure 3). To provide appropriate standards to assist in identification, an *H. pluvialis* ketolase and an Arabidopsis β -carotene hydroxylase were separately introduced into the β -carotene-accumulating *E. coli* to produce echinenone (3-keto- β , β -carotene) and canthaxanthin (3,3'-diketo- β , β -carotene) or β -cryptoxanthin (4-hydroxy- β , β -carotene) and zeaxanthin (4,4'-dihydroxy- β , β -carotene). None of the compounds formed in the presence of the ketolases of the invention (no difference was observed in products formed in the presence of the two different nucleic acid sequences of the invention) both migrate in the TLC system and have the absorption spectrum expected for echinenone, canthaxanthin, β -cryptoxanthin, or zeaxanthin. Two of the colored TLC bands produced in the presence of the Adonis ketolase cDNAs are orange in color. Orange band #1 has an absorption spectrum similar to that of canthaxanthin (see Figure 4) but migrates in a position that indicates a polarity intermediate to echinenone and β -carotene. Orange band #2 also has an absorption spectrum like that of canthaxanthin but migrates in a position that indicates a polarity intermediate to canthaxanthin and zeaxanthin (see Figure 3). The absorption spectra and TLC results suggest that the two orange products could be desaturated at the 3-4 positions of both rings (3,4,-didehydro; see Figure 2). Orange band #1 (see Figure 3) might then be 3,4,3',4'-tetrahydro- β , β -carotene. To substantially affect the absorption spectrum of the substrate β -carotene, any modifications very likely involve a carbon that lies in conjugation with the conjugated chain of carbon-carbon double bonds that constitute the chromophore (Goodwin, 1980; The Biochemistry of the Carotenoids, volume I; 2nd edition, Chapman and Hall). For the spectra obtained, only the carbons at the number 4 position of the two rings appear to be plausible locations for modification. The multitude and TLC migrations of the yellow and orange products produced from the symmetrical β -carotene, however, also indicates that the enzymes of the invention carry out more than a single type of reaction. The apparent homology of the ketolases of the invention to the Arabidopsis β -carotene hydroxylase would suggest that compounds with a hydroxyl at the 3 and/or 4 positions of one or both rings are another possible outcome (see Figure 2). In fact, such compounds have been identified in Adonis (see

above), and it has long been conjectured that a hydroxyl at position 4 is an intermediate in the formation of the 4-keto (e.g. crustaxanthin, a 3,3',4,4' tetrahydroxy carotenoid that might be a precursor for astaxanthin in the exoskeleton of the lobster). The histidine motifs and secondary structure in common to the hydroxylase and ketolase enzymes are characteristics of a large group of di-iron oxygenases whose members also include examples of desaturases (J. Shanklin, 1998, *Ann. Rev. Plant Physiol. Plant Mol. Biol.*), therefore a 3-4 desaturation (and/or perhaps a 2-3 desaturation in one or more of the yellow compounds) would also seem a plausible outcome.

To summarize the results of this example for the Adonis ketolases of the invention, a number of different carotenoids, including two with ketocarotenoid-like spectra, are produced from β -carotene via the action of the products of either of the two different nucleic acids of the invention. These orange compounds appear to be the major products. Truncation and fusion of the cDNAs to a stronger promoter in the vector pTrcHis (Invitrogen) was detrimental to growth of *E. coli* but did result in improved yield of the most polar orange product (orange band #2 in Figure 3). Introduction of a cyanobacterial ferredoxin did not change the yield or relative amounts of the various products. Without being bound by theory, it may be that the ketocarotenoids produced in flower petals of Adonis actually include the as yet unidentified orange compounds that are produced in *E. coli* using the nucleic acids of the invention.

EXAMPLE 2

Substrate specificity of the Adonis ketolases

Carotenoids with ϵ rings are common in plants. The ϵ ring differs from the b ring only in the position of the double bond within the ring (Figure 2). The ϵ ring is reported to be a poor substrate for the Arabidopsis b -carotene hydroxylase (Sun et al., 1996). The Adonis ketolase cDNAs were introduced into strains of *E. coli* engineered (Cunningham et al., 1996) to accumulate carotenoids with one or two ϵ rings (d -carotene and ϵ -carotene), or the acyclic carotenoid lycopene. TLC analysis of acetone extracts revealed that these carotenoids were not modified by the Adonis ketolases. as indicated by a lack of any new products formed. Products produced in *E. coli* engineered to accumulate zeaxanthin (Sun et al., 1996) appeared to be the same as

for β -carotene accumulating cultures indicating that a 3-OH is likely to be one of the functional groups introduced to the b ring by the Adonis ketolases. The more polar orange band produced from b-carotene through the action of the Adonis ketolases (e.g., orange band 2 in Figure 3), therefore, could very well be 3,3'-dihydroxy-3,4,3',4'-tetrahydro-b,b-carotene.

The references cited in the application, along with the following references, are incorporated by reference:

- Bouvier F, et al. (1998) Xanthophyll biosynthesis: molecular and functional characterization of carotenoid hydroxylases from pepper fruits (*Capsicum annuum* L.). *Biochim Biophys Acta*. 1391:320-8
- Breitenbach J, et al. (1996) Expression in *Escherichia coli* and properties of the carotene ketolase from *Haematococcus pluvialis*. *FEMS Microbiol Lett*. 140:241-6
- Cunningham FX Jr, Gantt E (1998) Genes and enzymes of carotenoid biosynthesis in plants. *Ann Rev Plant Physiol Plant Mol Biol* 49: 557-583
- 15 Fernandez-Gonzalez B, et al. (1997) A new type of asymmetrically acting beta-carotene ketolase is required for the synthesis of echinenone in the cyanobacterium *Synechocystis* sp. PCC 6803. *J Biol Chem*. 272:9728-33
- Fraser PD, et al. (1997) In vitro characterization of astaxanthin biosynthetic enzymes. *J Biol Chem*. 1997272:6128-35
- 20 Fraser PD, et al. (1998) Enzymic confirmation of reactions involved in routes to astaxanthin formation, elucidated using a direct substrate in vitro assay. *Eur J Biochem*. 252:229-36
- Harker M, et al. (1997) Biosynthesis of ketocarotenoids in transgenic cyanobacteria expressing the algal gene for beta-C-4-oxygenase, crtO. *FEBS Lett*. 404:129-34

- Kajiwara S, et al. (1995) Isolation and functional identification of a novel cDNA for astaxanthin biosynthesis from *Haematococcus pluvialis*, and astaxanthin synthesis in *Escherichia coli*. *Plant Mol Biol*. 29:343-52
- Lotan T, et al. (1995) Cloning and expression in *Escherichia coli* of the gene encoding
5 beta-C-4-oxygenase, that converts beta-carotene to the ketocarotenoid canthaxanthin
in *Haematococcus pluvialis*. *FEBS Lett*. 364:125-8
- Misawa N, et al. (1995) Canthaxanthin biosynthesis by the conversion of methylene to
keto groups in a hydrocarbon beta-carotene by a single gene. *Biochem Biophys Res*
10 *Commun*.209:867-76
- Misawa N, et al. (1995) Structure and functional analysis of a marine bacterial
carotenoid biosynthesis gene cluster and astaxanthin biosynthetic pathway proposed
at the gene level. *J Bacteriol*. 177:6575-84
- Miura Y, et al. (1998) Production of the carotenoids lycopene, beta-carotene, and
15 astaxanthin in the food yeast *Candida utilis*. *Appl Environ Microbiol*. 64:1226-9
- Shanklin J, et al. (1997) Mossbauer studies of alkane omega-hydroxylase: evidence for
a diiron cluster in an integral-membrane enzyme. *Proc Natl Acad Sci U S A*. 94:2981-6
- Shanklin J, Cahoon EB (1998) Desaturation and related modifications of fatty acids.
Ann Rev Plant Physiol Plant Mol Biol 49: 611-641
- 20 Wang CW, et al. Engineered isoprenoid pathway enhances astaxanthin production in
Escherichia coli. *Biotechnol Bioeng*. 1999 Jan 20;62(2):235-41.

I claim:

1. A method of producing a ketocarotenoid in a host cell, the method comprising inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the
5 nucleic acid sequence of SEQ ID NO: 1 or 3, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.
2. The method of claim 1, wherein the host cell is selected from the group
10 consisting of a bacterial cell, an algal cell and a plant cell.
3. A method of producing a ketocarotenoid in a host cell, the method comprising inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the
15 heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.
4. The method of claim 3, wherein the host cell is selected from the group consisting of a bacterial cell, an algal cell and a plant cell.
- 20 5. A method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method comprising
inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3,
25 wherein the heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed

host cell.

6. The method of claim 5, wherein the host cell is selected from the group consisting of a bacterial cell, an algal cell and a plant cell.

7. A method of modifying the production of carotenoids in a host cell, relative to an
5 untransformed host cell, the method comprising

inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably
10 linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

8. The method of claim 7, wherein the host cell is selected from the group
15 consisting of a bacterial cell, an algal cell and a plant cell.

9. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1.

10. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 3.

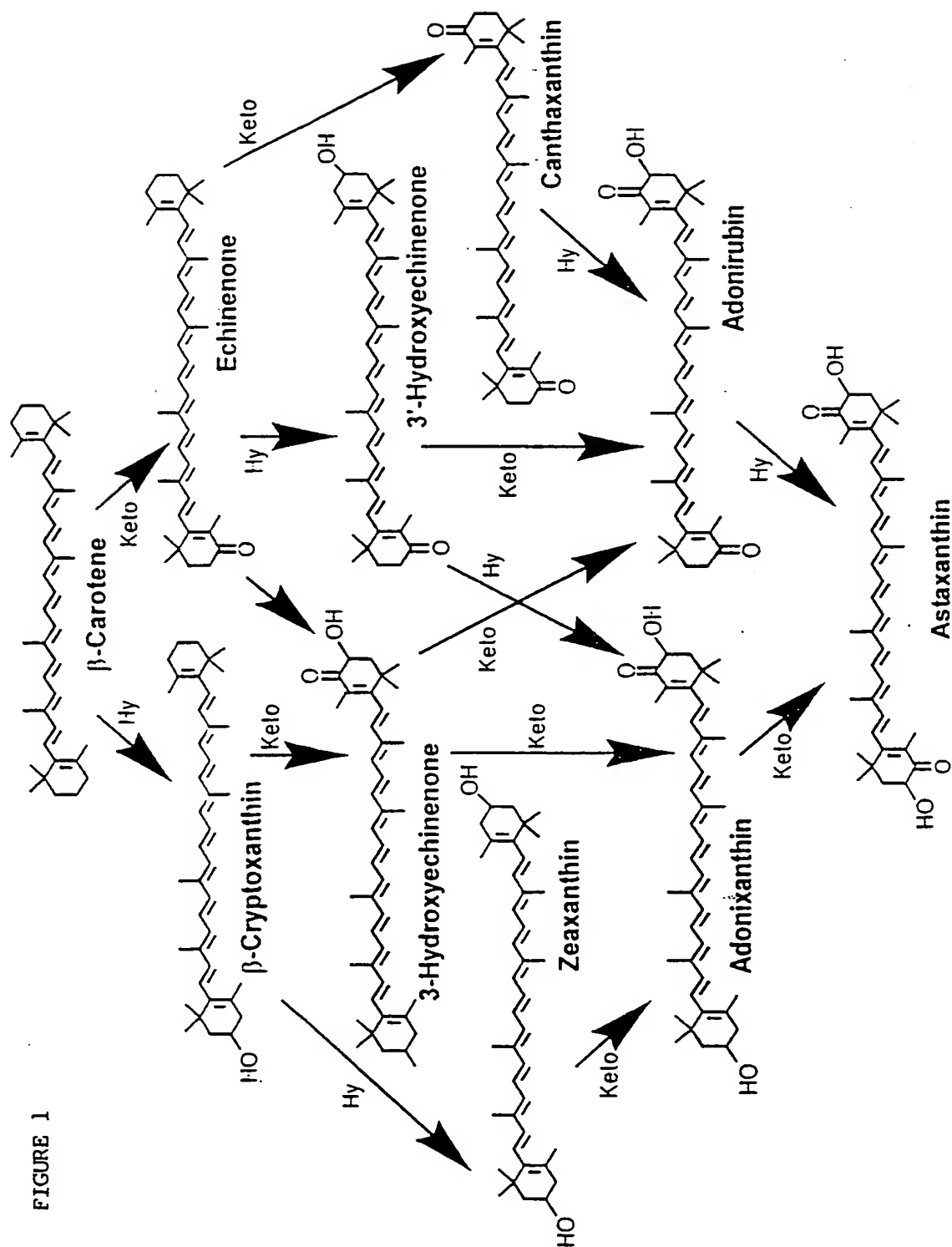
20 11. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2.

12. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ
25 ID NO: 4.

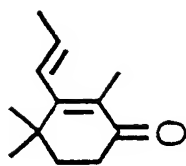
- 18 -

13. A vector which comprises the nucleic acid sequence of any one of claims 9-12, wherein the nucleic acid sequence is operably linked to a promoter.
14. A host cell which is transformed with the vector of claim 13.
15. The host cell of claim 14, wherein the host cell is selected from the group
5 consisting of a bacterial cell, an algal cell and a plant cell.
16. The host cell of claim 14, wherein the host cell is a photosynthetic cell.
17. The host cell of claim 14, wherein the host cell contains a ketocarotenoid.
18. The host cell of claim 14, wherein the host cell contains modified levels of carotenoids, relative to an untransformed host cell.
- 10 19. A purified ketolase enzyme which is encoded by the amino acid sequence of SEQ ID NO: 2.
20. A purified ketolase enzyme which is encoded by the amino acid sequence of SEQ ID NO: 4.

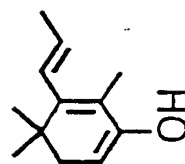
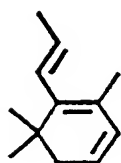
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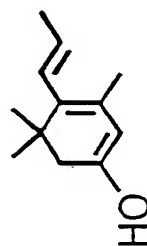
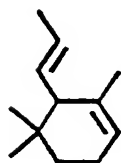
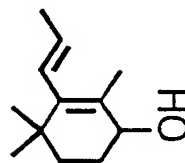
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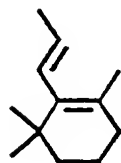
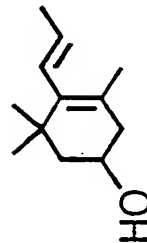
4-keto

3,4-dihydroxy
4-hydroxy

3,4-dihydro

3,4-dihydroxy
3-hydroxy ϵ ring

4-hydroxy

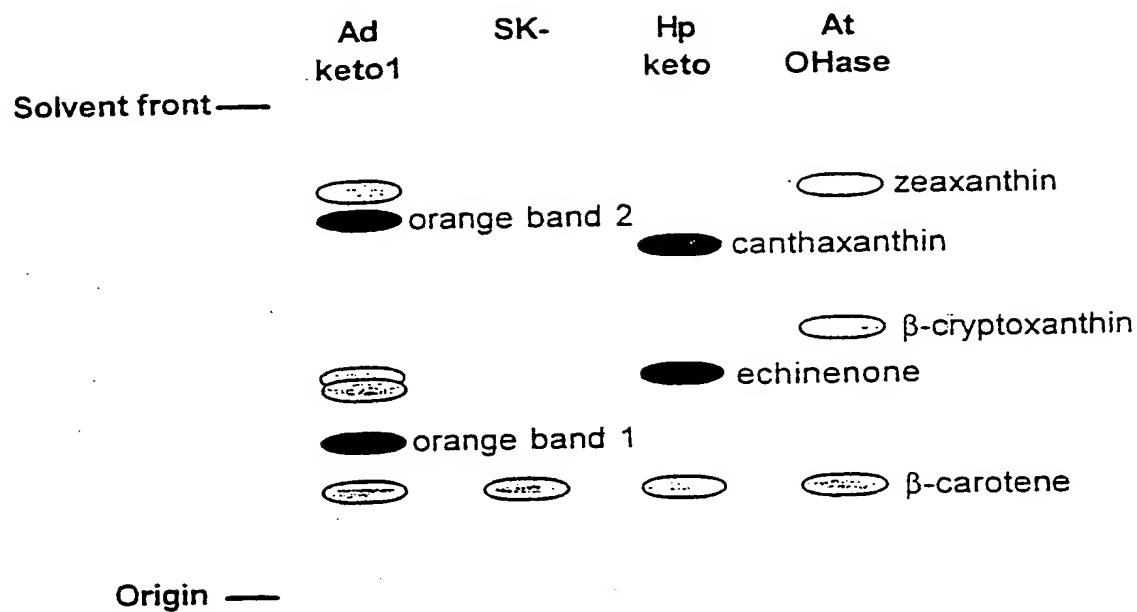
 β ring

3-hydroxy

FIGURE 2

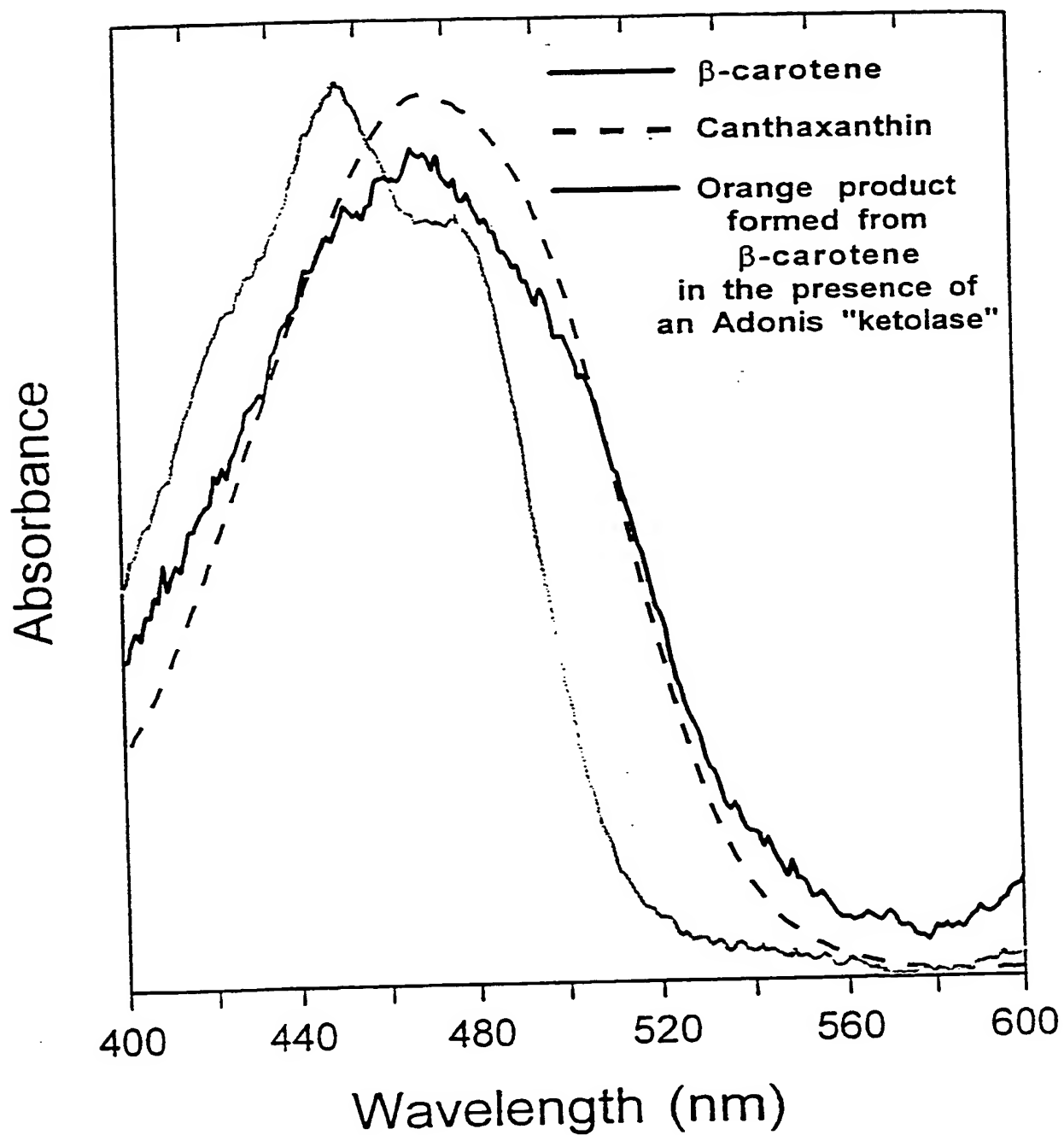
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FIGURE 3



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FIGURE 4



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Figure 6 [SEQ ID NO: 6]

MGLQEFGR
aisvfstsys fhknlllhsk qdilnrpcll fspvvvespm rkkkthraac
icsvaertrn ldipqieeee eneeelieqt dsgiihikkt lggkqsrrst
gsivapvscl gilsmigpav yfkfsrlmec gdipvaemgi tfaafvaaai
gteflsgwvnh kelwhdslwy ihkshhrsrk grfefndvfa iinalpaial
inygfsnegl lpgacfgtgltttvcgmayi flhnglshrr fpvglianvp
yfhklaaahq ihhsgkfqqv pfglflgpqe leevrggte iervisrtak
rtqsst*

Figure 7 [SEQ ID NO: 7]

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Figure 8 [SEQ ID NO: 8]

MGLQEFGR

aisvfssgys fyknilldsk pnilkppcll fspvvimspm rkkkkhgdpc
icsvagrtrn ldipqieeee enveelieqt dsdivhikkt lggkqskrpt
gsivapvscl gilsmigpav yfkfsrlmeg gdipvaemgi tfatfvaaav
gteflsawvh kelwheslwy ihkshhrsrk grfefndvfa iinalpaial
inygfsnegl lpgacfgvgl gttvcgmayi fihnglshrr fpvwlianvp
yfhklaaahq ihhsgkfqgv pfglflgpke leevrggte lervisrttk
rtqpst*

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Figure 9: Gap of SEQ ID NO: 9 and SEQ ID NO: 3

```
1 agcaatctcagtgttcagtacaagttattctttccacaagaatctcttgt 50
  ||||| ||||| ||||| || ||||| ||||| ||||| ||||| |||||
1 agcaatttcagtgttcaggttcagggtattctttctacaagaatctcttgt 50

51 tgcactcaaaacaagacattctcaaccgcccattgttgctcttctctcca 100
  || ||||| ||||| || ||||| ||||| ||||| ||||| |||||
51 tggactcaaaaccaaatattctcaaccccccatgcctgctattctctcca 100

101 gttgtgggtggagtcgcctatgagaaagaaaaagacacatcggtgctgcatg 150
  ||||| || ||||| ||||| ||||| ||||| ||||| ||||| |||||
101 gttgtgatcatgtcgcctatgagaaagaaaaagaaacatgggtgatccatg 150

151 tatctgctctgttgcagagagaacaaggaaccttgatattcctcaaattg 200
  ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
151 tatctgctccgttgcagggagaacaaggaaccttgatattcctcaaattg 200

201 aagaagaggaagagaacgaggaagaactaatagaacagacggattctggc 250
  ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
201 aagaagaggaagagaatgtggaagaactaatagaacagaccgattctgac 250

251 ataattcatataaagaaaacgctaggggggaaacaatcaagacgggtccac 300
  ||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
251 atagtgcataataaagaaaacactaggggggaaacaatcaaaacggccccac 300

301 tggctccattgtcgaccccgatatcttgtcttgggatcctttcaatgatcg 350
  ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
301 tggctccattgtcgaccccgatatcttgtcttgggatcctttcaatgattg 350
```

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Figure 9 (cont.)

351 gacctgctgtttacttcaagttttcacggctaataaggagtgtggagatatt 400
|||||
351 gacctgctgtttacttcaagttttcacggctaataaggagggtggagatata 400

401 cctgtcgcagaaatggggattacgtttgccgcctttgttgctgctgcat 450
|||||
401 cctgtagcagaaatggggattacgtttgccacctttgttgctgctgctgt 450

451 tggcacggaatttttgtcaggatgggttcacaaagaactctggcacgatt 500
|||||
451 tggcacggagtttttgtcagcatgggttcacaaagaactctggcacgagt 500

501 ctttgtggtacattcacaagtctcaccataggtcacgaaaaggccgcttc 550
|||||
501 ctttgtggtacattcacaagtctcaccatcggtcacgaaaaggccgcttc 550

551 gagttcaatgatgtgtttgctattattaacgcgcttcctgctattgctct 600
|||||
551 gagttcaatgatgtgtttgctattattaacgcgcttcctgctattgctct 600

601 tatcaattatggattctcaaataaggcctccttcctggagcctgctttg 650
|||||
601 tatcaattatggattctccaataaggcctccttcctggagcgtgctttg 650

651 gtaccggtccttggaaacgacagtctgtggcatggcttacatttttcttcac 700
||
651 gtgtcgggtccttggaaacaacagtctgtggatggcttacatttttcttcac 700

Figure 9 (cont.)

```

701 aatggcctttcacaccgaagggttcccagtagggcttattgcaaacgtccc 750
    ||||||| ||||||||||||||||||| ||||||| |||||||
701 aatggcctatcacaccgaagggttcccagtatggcttattgcaaacgtccc 750
    . . . . .
751 ttattttccacaagctggctgcagctcaccaaattccatcactcaggaaaat 800
    |||||||||||||||||||||||||||| || |||||||||||||||
751 ttattttccacaagctggctgcagctcaccaaatacaccactcaggaaaat 800
    . . . . .
801 ttcaggggtgtaccatttggcctgttccttggaccccaggaattggaagaa 850
    |||||||||||||||||||||||||||| |||||||||||||||
801 ttcaggggtgtaccatttggcctgttccttggaccaaggaattggaagaa 850
    . . . . .
851 gtaagaggaggcactgaagaattggagagggtgatcagtcgtacagctaa 900
    ||||||||||||||||||| ||||||||||| ||||||||||| ||||
851 gtaagaggaggcactgaagagttggagagggtaatcagtcgtacaactaa 900
    . . . . .
901 acgaacgcaatcatctacaTGAatcaactcttttacatttatgaggtttt 950
    ||||||||||| ||||||| ||||||| | ||||||| ||| |||||||
901 acgaacgcaaccatctaccTGAatcaatttttttacatatataagggtttt 950
    . . . . .
951 agtttatcggtgtta.caagtcacacatttgtgtcgttgtagtaattcaa 999
    ||||||||||||||| || ||||||| || ||||||| ||||| ||||
951 agtttatcggtgttataaaaatcacacatccgtatcgtttttagtaagtcaa 1000
    . . . . .
1000 agttaccatactcttttttagaatttttttttgatgtatagggtcgcgagg 1049
    ||||| ||||| || | | |||||||||||||||||||
1001 agttaagatacttccttcttagaataatttttttgatgtatagggtcgcggat 1050

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Figure 9 (cont.)

1050 ttacggttacaaaggccaaatctattggtgtggaattccattattaaaaa 1099

||| ||||| | | ||||| ||||| ||||| |||||

1051 atactgttac.....actattcggtgtggaattccattataaaaaa 1091

1100 taaaaattagagttttagtatttatctggtgatcaatatcaatatatt 1149

||| | |

1092 ataaaaaaaaaaaaaaaaaaaaa

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Figure 10: Gap of SEQ ID NO: 2 and SEQ ID NO: 4

1 AISVFSTSYSFHKNLLHLSKQDILNRPCLLFSPVVVESPMRKKKTHRAAC 50
|||||. |||:|||| || .|| |||||: ||||| | |
1 AISVFSSGYSFYKNLLDSKPNILKPPCLLFSPVVIMSPMRKKKKHGDPC 50
51 ICSVAERTRNLDIPQIEEEEEENEEELIEQTDSGIIHIKKTLLGGKQSRST 100
|||| |||||:|||||:| |
51 ICSVAGRTRNLDIPQIEEEEEENVEELIEQTDSDIVHIKKTLLGGKQSKRPT 100
101 GSIVAPVSCGLGILSMIGPAVYFKFSRLMECGDIPVAEMGITFAAFVAAAI 150
|||||:|||||:| |
101 GSIVAPVSCGLGILSMIGPAVYFKFSRLMEGGDIPVAEMGITFATFVAAAV 150
151 GTEFLSGWVHKELWHDLSLWYIHKSHHRSRKGRFEFNDVFAIINALPAIAL 200
||||| |||||:|||||:|||||:| |
151 GTEFLSAWVHKELWHESLWYIHKSHHRSRKGRFEFNDVFAIINALPAIAL 200
201 INYGFSNEGLLPACFGTGLGTTVCGMAYIFLHNGLSHRRFPVGLIANVP 250
|||||:|||||:|||||:| |
201 INYGFSNEGLLPACFGVGLGTTVCGMAYIFLHNGLSHRRFPVWLIANVP 250
251 YFHKLAAAHQIHHSKGKFQGVPFGLFLGPQELEEVRGGTEELERVISRTAK 300
|||||:|||||:| |
251 YFHKLAAAHQIHHSKGKFQGVPFGLFLGPKELEEVRGGTEELERVISRTTK 300
301 RTQSST* 307
||| |||
301 RTQPST* 307

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Figure 11: Gap of SEQ ID NO: 2 and Arabidopsis β -carotene hydroxylase (SEQ ID NO: 10)

```

1  AISVFSTSYSFHKNNLLHSHKQDILNRPCLLFSPVVVESPMRKKKTHRAAC 50
   .  ||. .|      ||      .  |  |  .      :  |  .
1  MAAXLSTAVTFKP...LHRSFSSSSSTDFRLRLPKSLSGFSPSLRFRFSV 47

51  ICSVAERTRNLDIPQIEEEEEENEEELIEQTDSGIIHIKKTLLGGKQSRRST 100
   |  || .|  |  |  |      :  :  :  .  |  |.  |||
48  CYVVEERRQNSPIENDERPESTSSTNAIDAAYLALRLAEKLERKKSERST 97

101 GSIVAPVSCLGILSMIGPAVYFKFSRLMECGDIPVAEMGITFAAFVAAAI 150
    |  | .|  ||  ||  |||::||  ||  |:|  .  ||  |||  |  ||:
98  YLIAAMLSSFGITSMAMVAVYYRFSWQMEGGEISMLEMFGTFALSVGA AV 147

151 GTEFLSGWVHKELWHDLSWYIHKSEHRSRKGRFEFNDVFALINALPAIAL 200
    |  || .  |  |:  |||  |||  .|.|||:  |.  ||  |||||:  ||  |||  |
148 GMEFWARWAHRALWHASLWNMHESHKPREGPFELNDVFALVNAGPAIGL 197

201 INYGFSNEGLLPACFGTGLGTTVCGMAYIFLHNGLSHRRFPVGLIANVP 250
    :.  |||  |.  ||.  ||  |||  |||  ||  |.  ||.  |.  ||  |:  ||||  ||.  ||
198 LSYGFFNKGLVPGLCFGAGLGITVFGIAYMFVHDGLVHKRFPVGPIADV 247

251 YFHKLAAAHQIHHSKGKFQGVFPGLFLGPQELEEVRGGTEELERVISRTAK 300
    |  |.  |||||:  ||.  ||  |||:  |||||  |.  ||||  ||  |||:  |||  |
248 YLRKVAAAHQLHHTDKFNGVPYGLFLGPKELEEV.GGNEELDKEISRRIK 296

301 RTQSST*..... 307
    .  .  .
297 SYKKASGSGSSSSS* 311

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Figure 12A (SEQ ID NO: 11)

1 CATACCATAA ATAGTAGAGG ACAACCTACA AACCAACCAC CAGAAACCTC 50
51 CAATGGCAGC

Figure 12B (SEQ ID NO: 12)

MAAAISVFSSGYSFYKNLLLD SKPNILKPPCLLFSPVVIMSPMRKKKKHGDPCICSVAGR
TRNLDPQIEEEEEENVEELIEQTDSDIVHEKKTLLGGKQSKRPTGSIVAPVSCLGILSMIG
PAVYFKFSRLMEGGDIPVAEMGITTFATFVAAAVGTEFLSAWVHKELWHESLWYIHKSHRR
SRKGRFEFNDVFAIINALPAIALINYGFSNEGLLPGACFGVGLGTTVCGMAYIFLHNGLS
HRRFPVWLIANVPYFHKLAAAHQIHHSKGKFGQVPPGLFLGPKELEEVRGGTEELERVISR
TTKRTQPST*

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Figure 13

```

      *      20      *      40      *      60      *
At1 : -----MAAXLSTAVTFKPLHRSFSSSSSTDFRLRLPKslagfspslR-----fkrfsvcyvve : 52
At2 : -----MAAGLSTIAVTTLKPLNRSSFSANHPistavfppslRFNGFRR-----rkiltvcfvve : 53
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Ca2 : TTGRYHYQLVWCQISFSSTSRYSYRHSPPFGPKPTPTTPSVYpitpfspnlGSILRCRR-----rpsitvcfvle : 71
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AdK6 : -----MAAAISVFSSGSHFYKNLLDSKPN LKPP lllfspvv MS MRKKKK-hgdp ic vag : 59
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      80      *      100      *      120      *      140
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At2 : erkqSSPMDDDNKPESTTSSEILMTS-----rlkklaekkkserftyliiaavmssfgit smaimavyvrrfs : 120
Ca1 : ddklyTAQSGKQSDTEAIGDEIFVETNEEKSLAVrlaekfarkkkserftyliiaavmssfgit smavisvvyvrrfs : 136
Ca2 : ddkfkTQFEAGREDIEMKIKEQISAT-----llaeklarckkserftyliiaavmssfgit smavmavyvrrfy : 137
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AdK6 : rrrnnddqqEEEEENVEER EQTDSDIV-----llkklgkksrrstt iiaavvs lgi smavmavyvrrfs : 128
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AdK6 : wcmeggripvaemgttfatva-aavgmefwarwahralwhaslwmbheshhkkp regpfelndvfainavpai : 201
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      *      *      *      *      *      *      *      *
      240      *      260      *      280      *
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At2 : gllyygffnkgllvpglcfagaglgltvfgmaymfvhdglvbkrfpvgsianvpylrkvaaahqhhhtdkfkgvpy : 268
Ca1 : affsfgnhkgllipglcfagaglgltvfgmaymfvhdglvbkrfpvgsiakvpyfqrvaaahqhhhtdkfkgvpy : 283
Ca2 : alldygffhkgllipglcfagaglgltvfgmaymfvhdglvbkrfpvgsianvpylrkvaaahqhhhtdkfngvpy : 284
AdK1 : alinygfsnkgllpglcfgtglgtvfgmayfahhglhbkrfpvgsianvpyfhk aaahqhhhtdkfkgvpy : 272
AdK6 : alinygfsnkgllpglcfvgglgtvfgmayfahhglhbkrfpvgsianvpyfhk aaahqhhhtdkfkgvpy : 275
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      *      *      *      *      *
      300      *      320      *
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At2 : glflgpkqeevvgGkaalekeisrriksykkaSGSSSSSS : 305
Ca1 : glflgpkaleevvg-ieelekevnrrriksikrl----- : 315
Ca2 : glflgpkaleevvg-leelekevnrrriksikrgs----- : 316
AdK1 : glflgpkaleevvgGtaalekeisrriksykkaSGSSSSSS : 306
AdK6 : glflgpkaleevvgGtaalekeisrriksykkaSGSSSSSS : 309
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<110> CUNNINGHAM, Francis X.

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USING THE GENES

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<140> Unknown

<141> 1999-05-21

<150> 60/086,460

<151> 1998-05-22

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Pro Val Val Val Glu Ser Pro Met Arg Lys Lys Lys Thr His Arg Ala
 35 40 45

Ala Cys Ile Cys Ser Val Ala Glu Arg Thr Arg Asn Leu Asp Ile Pro
 50 55 60

Gln Ile Glu Glu Glu Glu Glu Asn Glu Glu Glu Leu Ile Glu Gln Thr
 65 70 75 80

Asp Ser Gly Ile Ile His Ile Lys Lys Thr Leu Gly Gly Lys Gln Ser
 85 90 95

Arg Arg Ser Thr Gly Ser Ile Val Ala Pro Val Ser Cys Leu Gly Ile
 100 105 110

Leu Ser Met Ile Gly Pro Ala Val Tyr Phe Lys Phe Ser Arg Leu Met
 115 120 125

Glu Cys Gly Asp Ile Pro Val Ala Glu Met Gly Ile Thr Phe Ala Ala
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Phe Val Ala Ala Ala Ile Gly Thr Glu Phe Leu Ser Gly Trp Val His
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Lys Glu Leu Trp His Asp Ser Leu Trp Tyr Ile His Lys Ser His His
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Arg Ser Arg Lys Gly Arg Phe Glu Phe Asn Asp Val Phe Ala Ile Ile
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Asn Ala Leu Pro Ala Ile Ala Leu Ile Asn Tyr Gly Phe Ser Asn Glu
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Gly Leu Leu Pro Gly Ala Cys Phe Gly Thr Gly Leu Gly Thr Thr Val
 210 215 220

Cys Gly Met Ala Tyr Ile Phe Leu His Asn Gly Leu Ser His Arg Arg

225 230 235 240
 Phe Pro Val Gly Leu Ile Ala Asn Val Pro Tyr Phe His Lys Leu Ala
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 35 40 45
 Pro Cys Ile Cys Ser Val Ala Gly Arg Thr Arg Asn Leu Asp Ile Pro
 50 55 60
 Gln Ile Glu Glu Glu Glu Glu Asn Val Glu Glu Leu Ile Glu Gln Thr
 65 70 75 80
 Asp Ser Asp Ile Val His Ile Lys Lys Thr Leu Gly Gly Lys Gln Ser
 85 90 95
 Lys Arg Pro Thr Gly Ser Ile Val Ala Pro Val Ser Cys Leu Gly Ile
 100 105 110
 Leu Ser Met Ile Gly Pro Ala Val Tyr Phe Lys Phe Ser Arg Leu Met
 115 120 125
 Glu Gly Gly Asp Ile Pro Val Ala Glu Met Gly Ile Thr Phe Ala Thr
 130 135 140
 Phe Val Ala Ala Ala Val Gly Thr Glu Phe Leu Ser Ala Trp Val His
 145 150 155 160
 Lys Glu Leu Trp His Glu Ser Leu Trp Tyr Ile His Lys Ser His His
 165 170 175
 Arg Ser Arg Lys Gly Arg Phe Glu Phe Asn Asp Val Phe Ala Ile Ile
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 35 40 45

Met Arg Lys Lys Lys Thr His Arg Ala Ala Cys Ile Cys Ser Val Ala
 50 55 60

Glu Arg Thr Arg Asn Leu Asp Ile Pro Gln Ile Glu Glu Glu Glu Glu
 65 70 75 80

Asn Glu Glu Glu Leu Ile Glu Gln Thr Asp Ser Gly Ile Ile His Ile
 85 90 95

Lys Lys Thr Leu Gly Gly Lys Gln Ser Arg Arg Ser Thr Gly Ser Ile
 100 105 110

Val Ala Pro Val Ser Cys Leu Gly Ile Leu Ser Met Ile Gly Pro Ala
 115 120 125

Val Tyr Phe Lys Phe Ser Arg Leu Met Glu Cys Gly Asp Ile Pro Val
 130 135 140

Ala Glu Met Gly Ile Thr Phe Ala Ala Phe Val Ala Ala Ala Ile Gly
 145 150 155 160

Thr Glu Phe Leu Ser Gly Trp Val His Lys Glu Leu Trp His Asp Ser
 165 170 175

Leu Trp Tyr Ile His Lys Ser His His Arg Ser Arg Lys Gly Arg Phe
 180 185 190

Glu Phe Asn Asp Val Phe Ala Ile Ile Asn Ala Leu Pro Ala Ile Ala
 195 200 205

Leu Ile Asn Tyr Gly Phe Ser Asn Glu Gly Leu Leu Pro Gly Ala Cys
 210 215 220

Phe Gly Thr Gly Leu Gly Thr Thr Val Cys Gly Met Ala Tyr Ile Phe
 225 230 235 240

Leu His Asn Gly Leu Ser His Arg Arg Phe Pro Val Gly Leu Ile Ala
 245 250 255

Asn Val Pro Tyr Phe His Lys Leu Ala Ala Ala His Gln Ile His His
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Ser Gly Lys Phe Gln Gly Val Pro Phe Gly Leu Phe Leu Gly Pro Gln
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Leu Lys Pro Pro Cys Leu Leu Phe Ser Pro Val Val Ile Met Ser Pro	35	40	45
Met Arg Lys Lys Lys Lys His Gly Asp Pro Cys Ile Cys Ser Val Ala	50	55	60
Gly Arg Thr Arg Asn Leu Asp Ile Pro Gln Ile Glu Glu Glu Glu Glu	65	70	75
Asn Val Glu Glu Leu Ile Glu Gln Thr Asp Ser Asp Ile Val His Ile	85	90	95
Lys Lys Thr Leu Gly Gly Lys Gln Ser Lys Arg Pro Thr Gly Ser Ile	100	105	110
Val Ala Pro Val Ser Cys Leu Gly Ile Leu Ser Met Ile Gly Pro Ala	115	120	125
Val Tyr Phe Lys Phe Ser Arg Leu Met Glu Gly Gly Asp Ile Pro Val	130	135	140
Ala Glu Met Gly Ile Thr Phe Ala Thr Phe Val Ala Ala Ala Val Gly	145	150	155
Thr Glu Phe Leu Ser Ala Trp Val His Lys Glu Leu Trp His Glu Ser	165	170	175
Leu Trp Tyr Ile His Lys Ser His His Arg Ser Arg Lys Gly Arg Phe	180	185	190
Glu Phe Asn Asp Val Phe Ala Ile Ile Asn Ala Leu Pro Ala Ile Ala	195	200	205
Leu Ile Asn Tyr Gly Phe Ser Asn Glu Gly Leu Leu Pro Gly Ala Cys	210	215	220
Phe Gly Val Gly Leu Gly Thr Thr Val Cys Gly Met Ala Tyr Ile Phe	225	230	235
Leu His Asn Gly Leu Ser His Arg Arg Phe Pro Val Trp Leu Ile Ala	245	250	255
Asn Val Pro Tyr Phe His Lys Leu Ala Ala Ala His Gln Ile His His			

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 35 40 45

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 50 55 60

Arg Pro Glu Ser Thr Ser Ser Thr Asn Ala Ile Asp Ala Glu Tyr Leu
 65 70 75 80

Ala Leu Arg Leu Ala Glu Lys Leu Glu Arg Lys Lys Ser Glu Arg Ser
 85 90 95

Thr Tyr Leu Ile Ala Ala Met Leu Ser Ser Phe Gly Ile Thr Ser Met
 100 105 110

Ala Val Met Ala Val Tyr Tyr Arg Phe Ser Trp Gln Met Glu Gly Gly
 115 120 125

Glu Ile Ser Met Leu Glu Met Phe Gly Thr Phe Ala Leu Ser Val Gly
 130 135 140

Ala Ala Val Gly Met Glu Phe Trp Ala Arg Trp Ala His Arg Ala Leu
 145 150 155 160

Trp His Ala Ser Leu Trp Asn Met His Glu Ser His His Lys Pro Arg
 165 170 175

Glu Gly Pro Phe Glu Leu Asn Asp Val Phe Ala Ile Val Asn Ala Gly
 180 185 190

Pro Ala Ile Gly Leu Leu Ser Tyr Gly Phe Phe Asn Lys Gly Leu Val
 195 200 205

Pro Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Ile
 210 215 220

Ala Tyr Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val
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Gly Pro Ile Ala Asp Val Pro Tyr Leu Arg Lys Val Ala Ala Ala His
 245 250 255

Gln Leu His His Thr Asp Lys Phe Asn Gly Val Pro Tyr Gly Leu Phe
 260 265 270

Leu Gly Pro Lys Glu Leu Glu Glu Val Gly Gly Asn Glu Glu Leu Asp
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His Gly Asp Pro Cys Ile Cys Ser Val Ala Gly Arg Thr Arg Asn Leu
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Glu Gln Thr Asp Ser Asp Ile Val His Ile Lys Lys Thr Leu Gly Gly
 85 90 95

Lys Gln Ser Lys Arg Pro Thr Gly Ser Ile Val Ala Pro Val Ser Cys
 100 105 110

Leu Gly Ile Leu Ser Met Ile Gly Pro Ala Val Tyr Phe Lys Phe Ser
 115 120 125

Arg Leu Met Glu Gly Gly Asp Ile Pro Val Ala Glu Met Gly Ile Thr
 130 135 140
 Phe Ala Thr Phe Val Ala Ala Ala Val Gly Thr Glu Phe Leu Ser Ala
 145 150 155 160
 Trp Val His Lys Glu Leu Trp His Glu Ser Leu Trp Tyr Ile His Lys
 165 170 175
 Ser His His Arg Ser Arg Lys Gly Arg Phe Glu Phe Asn Asp Val Phe
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 Ala Ile Ile Asn Ala Leu Pro Ala Ile Ala Leu Ile Asn Tyr Gly Phe
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 Ser Asn Glu Gly Leu Leu Pro Gly Ala Cys Phe Gly Val Gly Leu Gly
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 Thr Thr Val Cys Gly Met Ala Tyr Ile Phe Leu His Asn Gly Leu Ser
 225 230 235 240
 His Arg Arg Phe Pro Val Trp Leu Ile Ala Asn Val Pro Tyr Phe His
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Cys Phe Val Val Glu Glu Arg Lys Gln Ser Ser Pro Met Asp Asp Asp
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Asn Lys Pro Glu Ser Thr Thr Ser Ser Ser Glu Ile Leu Met Thr Ser
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Val Val Gly Met Glu Phe Trp Ala Arg Trp Ala His Arg Ala Leu Trp
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Ala Ser Leu Trp His Met His Glu Ser His His Arg Pro Arg Glu Gly
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Ile Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Met Ala Tyr
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Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val Gly Pro
 245 250 255

Ile Ala Lys Val Pro Tyr Phe Gln Arg Val Ala Ala Ala His Gln Leu
 260 265 270

His His Ser Asp Lys Phe Asp Gly Val Pro Tyr Gly Leu Phe Leu Gly
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Val Pro Phe Ser Glu Met Phe Gly Thr Phe Ala Leu Ser Val Gly Ala
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Ala Val Gly Met Glu Phe Trp Ala Arg Trp Ala His Lys Ala Leu Trp
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His Ala Ser Leu Trp His Met His Glu Ser His His Lys Pro Arg Glu
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Gly Pro Phe Glu Leu Asn Asp Val Phe Ala Ile Ile Asn Ala Val Pro
 195 200 205

Ala Ile Ala Leu Leu Asp Tyr Gly Phe Phe His Lys Gly Leu Ile Pro
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Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Met Ala
 225 230 235 240

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/10455

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C12P 23/00, 7/26; C12N 9/02, 1/20, 15/00; C07H 21/04; C07K 14/00
US CL : 435/67, 148, 189, 252.3, 252.33, 320.1; 536/23.2, 23.6; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/67, 148, 189, 252.3, 252.33, 320.1; 536/23.2, 23.6; 530/350

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5,453,565 A (MAWSON) 26 September 1995, see abstract and claims.	1-20
A,E	US 5,910,433 A (KAJIWARA et al.) 08 June 1999, see the entire patent.	1-20
Y,P	US 5,811,273 A (MISAWA et al.) 22 September 1998, See abstract, column 30 - lines 48-58 and claims.	1-20

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

13 AUGUST 1999

Date of mailing of the international search report

29 OCT 1999

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/10455

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, STN Files: Medline, Caplus, Biosis, Agricola, Embase & Scisearch. Search terms used : beta carotene and ketolase, ketocarotenoid, Adonis aestivalis, carotenoid biosynthesis, gene? or dna or ma or nucleic acid? in various permutations and combinations.